

Neurobiology of social behavior abnormalities in autism and Williams syndrome

Boaz Barak^{1,2} & Guoping Feng¹⁻³

Social behavior is a basic behavior mediated by multiple brain regions and neural circuits, and is crucial for the survival and development of animals and humans. Two neuropsychiatric disorders that have prominent social behavior abnormalities are autism spectrum disorders (ASD), which is characterized mainly by hyposociability, and Williams syndrome (WS), whose subjects exhibit hypersociability. Here we review the unique properties of social behavior in ASD and WS, and discuss the major theories in social behavior in the context of these disorders. We conclude with a discussion of the research questions needing further exploration to enhance our understanding of social behavior abnormalities.

Introduction to social behavior

One of the most complicated behaviors humans and animals can perform is social behavior, which takes place between conspecifics and results in social relationships. Social behavior is based on the ability to properly communicate with others; individuals must sense, process and interpret social cues, as well as respond with appropriate behaviors. These functions are mediated by brain areas comprising the “social brain”¹, in particular, the medial prefrontal cortex (mPFC), amygdala, anterior insula, anterior cingulate cortex, inferior frontal gyrus and superior temporal sulcus (Fig. 1).

Two neuropsychiatric developmental disorders, ASD and WS, result in contrasting abnormalities in social behavior²: while ASD is characterized by social avoidance and lack of interest in social interactions, WS is characterized by uninhibited social interactions and overfriendliness. Although the opposing social behavior phenotypes of ASD and WS offer an opportunity to study neurobiological mechanisms of social abnormalities, the heterogeneity of ASD symptoms and genetics makes it complicated to directly compare the contrasted social behaviors. By contrast, the well-characterized genetic information of WS and its distinctive behavioral phenotype make the study of its neurogenetics more accessible and could help to understand the relationship among genes, neural circuitry, physiology and social behavior. In this review, we compare and contrast the symptoms, genetics and related clinical findings of these two disorders with the hope that further comparative studies will uncover underlying neurobiological mechanisms of social behavior abnormality.

Contrasting social behavior abnormalities in ASD and WS

Autism spectrum disorders. ASDs are a group of heterogeneous neurodevelopmental disorders characterized according to the Diagnostic

and Statistical Manual of Mental Disorders, Fifth Edition (DSM-5) by (i) deficits in social communication and social interaction and (ii) stereotyped, repetitive behavior³ with narrow restricted interests⁴, often accompanied by sensory abnormalities and language development delay or absence. These symptoms must be present in early childhood and impede the individual’s everyday activity. Autism, from the Greek words *autos* (“self”) and *ismos* (“action”), was described initially by Kanner in 1943 (ref. 5) as a congenital lack of interest in other people. Nowadays, ASD affects 1 in 68 children in the United States⁶ (but see ref. 7), with approximately five times as many boys affected as girls⁸.

ASD is one of the most heritable common psychiatric disorders, indicating that genetics are central to ASD etiology. Nevertheless, the genetic contribution to pathophysiology is challenging to explore because of incomplete penetrance, a large number of susceptibility genes, and complex gene–environment interactions. While genome-wide association studies have yet to yield replicable common variants for ASD, possibly owing to small sample sizes, studies of copy number variants and single nucleotide polymorphisms have provided gene candidates for further study^{9–14}. Many of the ASD-linked genes encode synaptic proteins¹⁵ at glutamatergic synapses (Fig. 2 and Supplementary Table 1), most of them acting postsynaptically¹⁶, indicating that excitatory synaptic dysfunction may be a key pathophysiology in ASD. However, our understanding of the molecular architecture of inhibitory synapses is very limited, so further studies on the basic biology of inhibitory synapses may shed new lights on etiology and pathology of ASD.

Deficient social behavior in ASDs. Although ASDs are heterogeneous in etiology and symptoms, a common central feature is social behavior deficit unrelated to cognitive dysfunction⁴. Part of the deficit includes impairments in social interaction, such as the inability to initiate social interactions or develop relationships, lack of social or emotional reciprocity, lack of interest in others’ emotions¹⁷, communication deficits including impaired speech development and poor expressive language¹⁸, impairment in nonverbal social interaction, and lack of interest in sharing enjoyment and interests with others¹⁹.

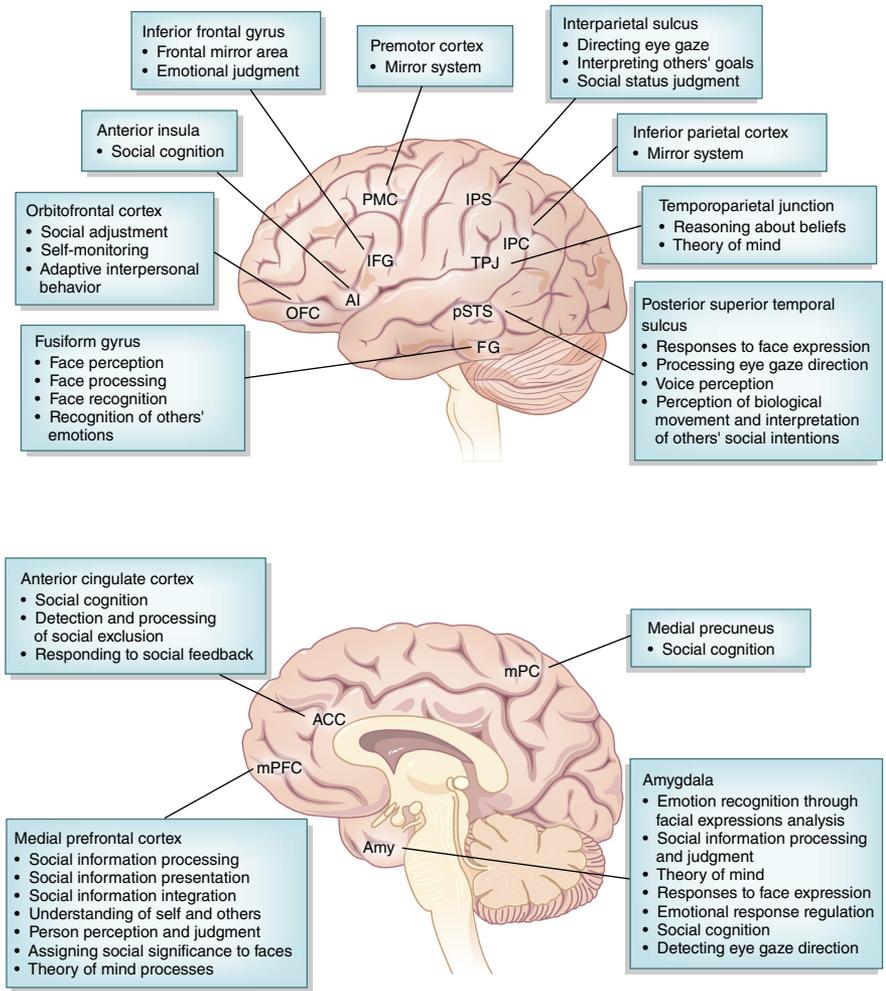
¹McGovern Institute for Brain Research, MIT, Cambridge, Massachusetts, USA.

²Department of Brain & Cognitive Sciences, MIT, Cambridge, Massachusetts, USA.

³Stanley Center for Psychiatric Research, Broad Institute of MIT and Harvard, Cambridge, Massachusetts, USA. Correspondence should be addressed to G.F. (feng@mit.edu).

Received 10 October 2015; accepted 22 February 2016; published online 26 April 2016; doi:10.1038/nrn.4276

Figure 1 Anatomical and functional brain areas related to social behavior. Brain regions of high relevancy to social behavior, participating in the different aspects of social behavior and their contribution to social behavior.



The earliest evidence of impaired social behavior that arises during the course of ASD is impaired selective attention and lack of innate preference of newborns for human voice²⁰ and face²¹ over other sounds and visual stimuli. Infants with ASD demonstrate impaired joint attention²², the ability to share eye gaze focus on an object following the alert of one individual to the other by pointing or gazing. In typical older children, the increased ability to communicate verbally with others results in more complex social behavior, including shared play and interactions with other children; these abilities, impaired in children with ASD²³, emphasize the profound differences between a typical child and one with ASD and are one of the major alerts for testing the child for ASD. These social behavior deficits continue in adults with ASDs, impairing their behavior.

Williams syndrome. WS or Williams-Beuren syndrome is a rare multisystemic neurodevelopmental genetic disorder named after John C.P. Williams, who was the first to describe the syndrome in 1961 (ref. 24). Physically, WS is associated with cardiovascular difficulties, growth abnormalities, connective tissue and endocrine abnormalities, and specific 'elfin' facial and physical anomalies. Mentally, WS is associated with distinctive central cognitive and personality profiles, independent of IQ, which include overfriendliness (frequently termed the "cocktail party personality"), increased empathy, mental retardation²⁵, strength in verbal and language skills²⁶, weaknesses in visual-spatial skills²⁷, increased musical interest and emotional reactivity to music²⁸ and elevated anxiety derived from fear and specific phobias²⁹.

WS prevalence is between 1 in 7,500 (ref. 30) and 1 in 20,000 (ref. 31) individuals, and is caused by a hemizygous deletion of about 25 genes at the 7q11.23 region on chromosome 7 (ref. 32). These genes are part of the WS chromosome region (WSCR), estimated to be about 1.6 megabases, the typical deletion in ~95% of subjects. The other ~5% of subjects have longer deletions of ~1.84 megabases^{33,34} or other extremely rare types of deletions³⁵⁻³⁷.

Interestingly, individuals with one or two extra copies of the WSCR genes due to WSCR duplication (Dup7) have an ASD-related phenotype characterized by developmental impairments, poor eye contact, anxiety disorder, repetitive behavior, hyposocial behavior and severe expressive language delay, which is the most commonly reported feature of Dup7 (refs. 38-42), although the range of these phenotypes is larger and much less studied than in WS. Overall, these phenotypes suggest that WSCR genes are dosage-dependent and may affect language skills and development.

Hypersociability in WS. Although WS is characterized by multiple physiological and mental features, the hypersociability phenotype is a

striking feature of WS and seemingly the opposite of the typical phenotype seen in ASDs. This unique social behavior is the reason why, in one of the first studies to characterize subjects with WS, they were described as individuals who "love everyone, are loved by everyone, and are very charming"⁴³.

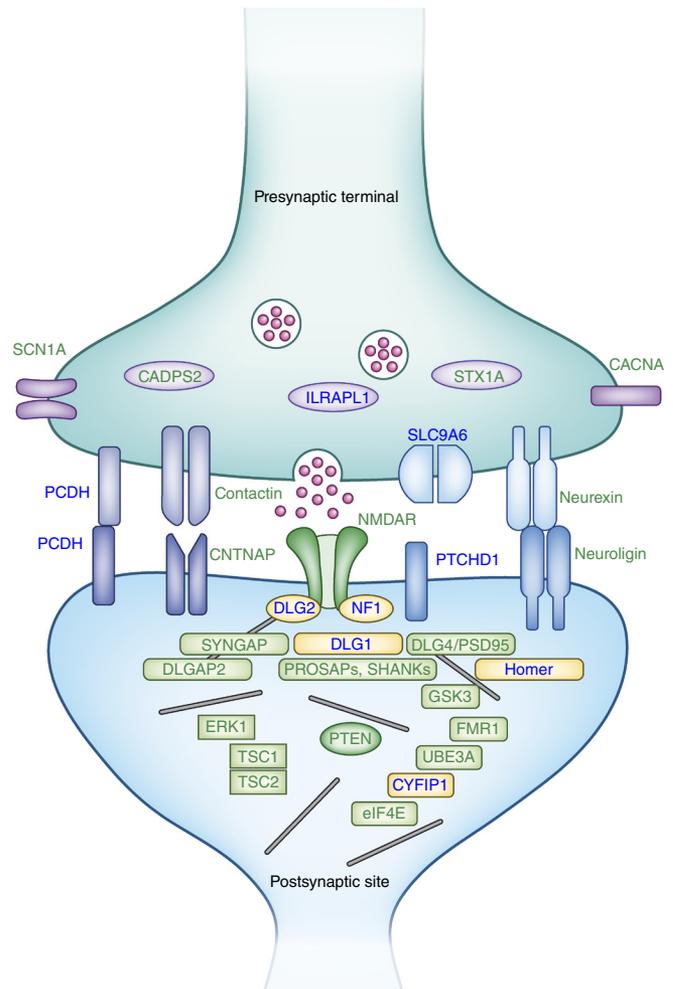
In WS, the gregarious personality is characterized by a consistent increased interest in and approach to strangers⁴⁴, overfriendliness that is positively correlated with age^{45,46}, and excessive empathy but poor social judgment ability. One of the main reasons suggested for the hypersociability in WS is the substantial attention bias toward any kind of social stimuli, with a special interest in human faces⁴⁷ (but see ref. 48), in contrast to the behavior seen in subjects with ASDs.

The distinctive intense gazing pattern begins at infancy and continues throughout development⁴⁹. While processing faces, individuals with WS demonstrate atypical patterns, with increased focus on faces and eyes⁴⁷ that lasts longer than in typically developed controls⁵⁰.

Toddlers and young children with WS continue showing higher sociability behavior, as measured by parental ratings of their child's social behavior⁵¹, and by their high engagement in dyadic, face-to-face interactions compared to control children⁵². Hypersociability persists in older children⁴⁵ and into adulthood, in which a longitudinal study found improved yet still abnormal social and adaptive functioning⁵³.

Another difficulty for subjects with WS is accurate perception of emotions. In particular, individuals with WS demonstrate difficulties in detecting social fear signals given through facial expressions and voices⁵⁴ and show less arousal in response to angry faces⁵⁵ than

Figure 2 Synaptic proteins at the glutamatergic synapse encoded by ASD-related genes. These synaptic proteins participate in the formation, stabilization and function of the synapse. Proteins in which a mutation in their encoding genes affects social behavior are marked in green, while those with no direct evidence are marked in blue. Presynaptic proteins are SCN1A (sodium channel protein type 1), CADPS2 (calcium-dependent secretion activator 2), ILRAPL1 (interleukin-1 receptor accessory protein-like 1), STX1A (syntaxin-1A), CACNA (voltage-dependent calcium channel subunit- α), PCDH (protocadherin), contactin, SLC9A6 (sodium-hydrogen exchanger) and neuexin; postsynaptic proteins are PCDH, CNTNAP (contactin-associated protein), NMDAR (glutamate receptor), PTCHD1 (patched domain-containing protein 1), neuroligin, DLG2 (Disks large homolog 2), NF1 (neurofibromin 1), PROSAPs (SH3 domain and ankyrin repeat containing proteins) and SHANKs (proline-rich synapse-associated proteins), homer, GSK3 (glycogen synthase kinase-3), ERK1 (mitogen-activated protein kinase 3), TSC1 (tuberous sclerosis 1), TSC2 (tuberous sclerosis 2), PTEN (phosphatase and tensin homolog), FMR1 (fragile X mental retardation protein), UBE3A (ubiquitin protein ligase E3A), CYFIP1 (cytoplasmic FMR1-interacting protein-1), and eIF4E (eukaryotic translation initiation factor 4E).



non-impaired controls. Individuals with WS also tend to have greater attention bias for positive than negative facial expression⁵⁶, and they rate happy faces⁵⁷ and unfamiliar faces⁵⁸ as more approachable than do control subjects.

A key factor that affects the cognitive phenotype in individuals with WS is the location of the shorter atypical microdeletions. Studies have found a classic behavioral and neurodevelopmental phenotype in cases where the atypical deletion includes the usual telomeric breakpoint, which results in deletion of the genes general transcription factor 2I (*Gtf2i*)⁵⁹ and *Gtf2i* repeat domain containing 1 (*Gtf2ird1*) from the general transcription factor 2I gene family^{60,61}. But, in cases where *Gtf2i* and *Gtf2ird1* genes are not deleted, only a mild behavioral and neurodevelopmental phenotype was found^{37,62,63}, suggesting that *Gtf2i* and *Gtf2ird1* deletion is important in the etiology of the behavioral and neurodevelopmental phenotype of WS.

Gtf2i encodes transcription factor II-I (TFII-I), a highly conserved and ubiquitously expressed multifunctional transcription factor that contains DNA-binding-I repeat domains, a leucine zipper and a nuclear localization signal⁶⁴. TFII-I regulates gene expression through interactions with tissue-specific transcription factors and complexes related to chromatin remodeling⁶⁵. Most WSCR deletions include both genes because *Gtf2i* and *Gtf2ird1* genes are in close proximity to each other; however, *Gtf2i* deletion has been shown to be more important for the WS social behavior phenotype. For example, by comparing the social behavior phenotype in rare cases of microdeletions sparing *Gtf2i* to those with the full WSCR deletion, Dai *et al.*⁶⁶ found the behavioral phenotype of the patient with the spared *Gtf2i* to be less social. Similarly, in individuals with different microdeletions sparing *Gtf2i*, Morris *et al.*⁶² found a WS cognitive profile but no mental retardation or intellectual difficulties. *Gtf2i* was also suggested to be highly involved in other neurobehavioral impairments of subjects with WS³⁵. In contrast, a patient with haploinsufficiency for *Gtf2ird1* but normal *Gtf2i* expression levels demonstrated normal social behavior but a delay in language acquisition⁶⁷.

In mice, homozygous deletion of *Gtf2i* causes embryonic lethality and severe developmental impairments⁶⁸, including neural tube defects and exencephaly. Heterozygous deletion of *Gtf2i* in mice results in impaired social habituation to an unfamiliar mouse, leading to increased time spent investigating the unfamiliar mouse as compared to that in wild-type mice⁶⁹. In a three-chamber social interaction and recognition test, *Gtf2i* heterozygous mice demonstrated about 50% higher preference ratio for interacting with an unfamiliar mouse than a novel object, compared to wild-type mice⁶⁹.

It is not known how transcriptional dysregulation resulting from *Gtf2i* deletion can lead to the hypersocial phenotype in WS, and there is no clear overlap in transcriptional dysregulations between WS and ASD. A recent study using induced pluripotent stem cells found that in the pluripotent state *Gtf2i* is already responsible for 10–20% of the transcriptional dysregulation in disease-relevant pathways in WS and Dup7 (ref. 70). It is therefore possible that transcriptional dysregulation as a result of *Gtf2i* deletion could result in impaired development of neural circuits that are crucial for normal social behavior from the very earliest development stages.

Etiology of social behavior abnormalities

Although the etiology of social behavior abnormalities in ASD and WS is still unclear, researchers have identified many associated anatomical and physiological changes. Because of the limitations inherent in studying human subjects, basic molecular and cellular research in animal models is crucial to better understand mechanisms underlying social behavior. Indeed, findings from animal studies have led to the development of several theories that relate to social behavior. However, humans and animals have evolved under different evolutionary pressures. Because of this evolutionary divergence, while molecular and cellular functions are largely comparable, social behaviors are much harder to compare. This is due to differences between animals and humans in the complexity of social behaviors, as well as the underlying motivations. Moreover, the sensory cues that lead to social response in these two groups

are substantially different and hence rely on the proper function of different neural circuits.

We will focus on three key theories, representing the physiological, functional, and systemic aspects of the theories in the field of social behavior. Since social behavior has been highly studied in the frame of ASD, these theories relate mainly to ASD rather than WS.

Social cognition in human studies. To properly perform social behavior, an individual needs to acquire, process, store and use social input from the environment to decide on and take proper social actions, the sum of which is called social cognition. Social cognition also relates to the process of understanding others or one's own thoughts, mental states and feelings ("theory of mind," or mentalization)⁷¹. This process is impaired in children with ASD⁷² and may result in impaired social information analysis and abnormal responses⁷³. These functions involve mainly the functionality of cortical brain regions (Fig. 1). Hence, cortical dysfunction might lead to cognitive dysfunction in general, and specifically to impairments in social cognition and sensory integration. Importantly, it is still unknown why social cognition is specifically impaired in subjects with otherwise normal cognition.

Cortical dysfunction can be the result of improper development of the cortex; in early stages of development, genes determine and regulate the formation of the brain, including its cells, synapses and neural circuits. However, later the complex interaction between genes and the subject's environment may lead to alterations in brain development that will result in an inability to respond to the environment⁷⁴. Genetic mutations can lead, for example, to improper synapse formation or imbalanced cellular activity between GABAergic and glutamatergic neurons. This may result in lack of proper development and function of inhibitory circuits that are essential for balanced neural activity during critical periods, and similar development and functional issues in brain regions and circuits essential for social behavior⁷⁵. A lack of early experience-dependent development may result in impaired development of primary sensory circuits, for example, which could lead to further impairments in more complex functions, governed by higher-order neural circuits that develop later. Consequently, the social brain does not receive proper stimulation and experience with integrating and processing social-related inputs, nor with the execution of social decisions and actions, leading to social disabilities.

Following this logic, and focusing for example on the need to properly process social information, improper function of cortical and subcortical brain regions results in sensory integration and multisensory processing problems, and indeed, sensory abnormalities are found in 90% of children with ASD⁷⁶. Not properly integrating and processing the social information around them, overstimulated subjects may have difficulties in changing their attention to social-related information, resulting in improper social orientation that causes behavioral deficits. Overwhelmed by stimuli, subjects with ASD might therefore tend to perform repetitive movements that return them to their 'safe zone' and relieve their anxiety.

Because most ASD and WS research focuses on subjects of toddler age and older, prenatal and early postnatal processes responsible for early development deficits are less understood (for review, see ref. 77). Consequently, it is difficult to differentiate between causes and effects: that is, whether a primary disruption of brain development leads to social abnormalities or whether an improper interaction with the environment leads to undeveloped social-related brain regions. Thus, more research needs to be done during infancy and followed up in a longitudinal manner, as this will also enable earlier diagnosis, earlier intervention, and identification of earlier-acting mechanisms.

This can be addressed by studying infants at high familial risk for ASD as part of prospective longitudinal studies⁷⁸.

A recent longitudinal magnetic resonance imaging (MRI) study examined the morphology of the corpus callosum in infants at high risk for ASD, as compared to low-risk controls. The findings from this study showed significantly increased corpus callosum area and thickness in children who were later diagnosed with ASD spectrum disorder starting at 6 months of age⁷⁹. An additional longitudinal MRI study on the development of white matter pathways in infants at high-risk for ASD found higher fractional anisotropy in 6-month-old subjects with ASD, followed by blunted developmental trajectories, resulting in lower fractional anisotropy by 24 months (ref. 80). Another study suggested that an increased cortical surface area, resulting from an increased rate of brain growth before age 2, is responsible for the brain enlargement in children with ASD⁸¹. More specifically, this enlargement in ASD toddlers is attributed to a generalized cerebral cortical enlargement, with an excessive temporal lobe white matter enlargement⁸¹. Yet another longitudinal MRI study also found cerebral enlargement in ASD toddlers, including both gray and white matter, with the highest degree of enlargement in frontal, temporal and cingulate cortices⁸². Interestingly, a different study on 6-month-old infants at high risk and their low-risk controls did not find significant differences in intracranial, cerebrum, cerebellum or lateral ventricle volume or head circumference⁸³. Additionally, young boys with ASD had decreased volumes of white matter and the dorso-lateral region of the frontal cortex as compared with control subjects, suggesting delayed development of these regions⁸⁴.

Imaging studies in adult ASD patients support changes particularly in mPFC. An MRI study found that subjects with ASD have decreased mPFC activation during mentalizing and weaker functional connectivity of the mPFC to other brain regions, as compared to control subjects⁸⁵. These findings suggest that subjects with ASD use different neural circuits and patterns of activation than control subjects to analyze their own and other people's emotions. Another study demonstrated that the mPFC is also involved in joint attention in subjects with ASD⁸⁶: it found a lack of signal differentiation and atypical pattern of dorsal mPFC activation in subjects with ASD compared to control subjects during a task that required joint attention. Lastly, studies have demonstrated abnormal local connectivity^{87,88} as well as abnormal long-range connectivity in ASD subjects, with the latter linked to altered development of white matter in multiple brain regions (for review, see ref. 89). However, the cellular mechanisms underlying these axonal disorganizations are not fully known.

In support of the imaging findings, histological examination of the frontal cortices of subjects with ASD has found abnormal neuronal morphology⁹⁰ and reduced minicolumns⁹¹, suggesting that improper development of this cortical area might play a role in impaired social input integration.

Frontal lobe dysfunction is also related to the WS hypersociability profile, as those regions have a role in regulating and suppressing actions that are socially inappropriate. The relatively low intelligence of patients with WS presents a challenge when comparing cortical function between subjects with WS and their control groups; it is important to select experimental and control subjects with comparable levels of intelligence. Examining subjects with WS who had normal intelligence, Meyer-Lindenberg *et al.* showed abnormal activity of the prefrontal cortex, including the orbitofrontal cortex (OFC), as a function of task, as compared to normal controls⁹². Additionally, Meyer-Lindenberg *et al.* found relatively reduced task-based connectivity between OFC and the amygdala in subjects with WS compared to controls⁹². Functionally, lesions of the OFC were associated with

social disinhibition, suggesting that abnormal OFC activity in subjects with WS might be responsible for the disinhibition of social approach. Deficits in regulating actions were suggested to be responsible for the high social approach behaviors of subjects with WS, resulting in poor social response inhibition due to frontal lobe dysfunction^{93,94}. Indeed, Porter *et al.* showed similarities in social approach suppression in subjects with WS and those with frontal lobe damage⁹⁴. Both types of subjects express impulsive social approach behavior and verbalize inappropriate thoughts, likely as a result of poor response inhibition⁹⁴. This was also noted in a recent study in children with WS, which demonstrated that frontal lobe–controlled response inhibition capability is the strongest indicator of social approach behavior⁹⁵. Lastly, abnormal cortical activity in subjects with WS was observed in the right OFC, showing an opposite pattern of OFC activation in response to positive and negative emotional faces⁹⁶. Moreover, this same study showed reduced activation of the right amygdala in response to negative faces as compared to that in typically developing controls⁹⁶.

Cortical dysfunction revealed by animal studies. Although the neurophysiological substrates for social behavior abnormalities are unknown, on the basis of human and animal model studies we may speculate that excitatory-inhibitory (E/I) neuronal activity imbalance might explain the physiological mechanism of social behavior abnormalities⁹⁷. Changes in the E/I balance can result in hyper-⁹⁸ or hypoactivation⁹⁹ of specific brain regions and lead to dysfunction of the affected brain regions. For example, elevated excitatory activity specifically in mouse mPFC results in impaired social behavior¹⁰⁰, and, consistent with the E/I imbalance theory, elevated activation of inhibitory cells rescues the social deficits¹⁰⁰.

On a genetic level, the association of genes with social behavior is not straightforward, despite multiple animal models showing synaptic or circuit dysfunction accompanied by social behavior abnormalities. For instance, E/I imbalance can occur in cortical regions as a result of mutations in synaptic proteins such as Shanks^{98,101}, a family of key postsynaptic density (PSD) proteins located in glutamatergic synapses that, together with other postsynaptic proteins (SAPAP and PSD-95), forms a postsynaptic scaffolding complex (Fig. 2)^{102–104}. While ASD is considered a polygenic disorder in most cases, recent studies showed that genetic disruption of *Shank2* and *Shank3* in mice results in substantial physiological and biochemical alterations at synapses that may contribute to impaired social behavior^{99,105–110}.

The importance of E/I balance in the cortex was also demonstrated in a mouse model of Rett syndrome¹¹¹. Methyl-CpG-binding protein 2 (MeCP2) regulates the expression of many genes by acting as a transcriptional activator and repressor, and mutations in *Mecp2* are known as the primary cause of Rett syndrome. Specific deletion of *Mecp2* from either all GABAergic neurons in the nervous system (using *Viaat-Cre* mice) or a specific subset of GABAergic neurons in the forebrain (using *Dlx5/6-Cre* mice) resulted in mice with features of Rett syndrome and ASD (Supplementary Table 1)¹¹². Deletion of *Mecp2* resulted in a reduced inhibitory quantal size, demonstrating that specific disruption of inhibitory signaling is sufficient to recapitulate ASD behaviors¹¹².

Social cognition relies on proper sensing and integration of sensory and social input, and indeed, sensory abnormalities are common in ASD. Recently, two studies on mouse models of ASD demonstrated the importance of the inhibitory system in sensory input processing and integration. Impaired maturation of the inhibitory system in the insula cortex of BTBR mice results in decreased inhibitory neurotransmission and increased excitatory neurotransmission, affecting multisensory integration⁹⁸. Treatment with a benzodiazepine, a

positive modulator of GABAergic transmission, rescues the impairment when provided early in postnatal development, but not when provided at a later age⁹⁸. Furthermore, the GABA-B agonist (R)-baclofen has also been shown to reverse social deficits in BTBR mice¹¹³. In another study, impaired function of the inhibitory system affected sensory input processing in the somatosensory barrel cortex of juvenile mice with an R451C substitution in Nlgn3 (neuroligin 3)¹¹⁴, a postsynaptic protein important for trans-synaptic cell adhesion (Fig. 2). Cellot *et al.* recently showed that R451C mutation affects the probability of GABA release from parvalbumin-expressing interneurons, impairing their modulation of principal cells in layer IV of the somatosensory barrel cortex¹¹⁴. This leads to a shift in E/I balance and affects the generation of cortical gamma rhythms associated with high cognitive functions such as social behavior.

Currently, neurobiological knowledge of the role of synaptic signaling in WS is extremely limited. Therefore, it would be of great interest to study the developmental abnormalities at the molecular and cellular levels that lead to cortical dysfunction in WS.

The amygdala theory. The amygdala, an almond-shaped region comprising at least 13 nuclei with unique functions, is part of the limbic system. The amygdala, which is highly connected to brain regions responsible for sensory input and autonomic systems, takes part in central functions and processes that are crucial for proper social behavior and emotional processing, and hence is suggested as a central component of the social brain (Fig. 1). The amygdala's roles in social behavior include the processing of emotional reactions, memories and visual social stimuli; creation and control of anxiety; and recognizing social emotion from faces. The amygdala also has a central role in the recognition of faces and facial emotion, and in mediating eye gaze¹¹⁵, such that subjects with complete amygdala lesions, like those with ASD, show impaired eye contact¹¹⁶. In high-functioning subjects with ASD, an impaired ability is found in recognizing social information from faces, as in subjects with focal bilateral amygdala damage¹¹⁷. Additionally, impaired social judgment was demonstrated in subjects with amygdala lesions¹¹⁸; conversely, deep-brain stimulation of the amygdala improved social behavior in a boy with ASD¹¹⁹.

Anatomically, children with ASD have larger right and left amygdala volumes than those without, although this difference is gone by adolescence¹²⁰. An increased amygdala volume was found also in subjects with WS^{121,122}, together with a positive correlation between right amygdala volume and the approachability of faces¹²². These findings support the notion that abnormalities in amygdala development and function may contribute to deficits in social judgment, emotional information processing and face expression perception, leading to abnormal emotional reactions and social behavior abnormalities in ASD and WS. Current knowledge is still contradictory, and the opposing social behaviors seen in ASD and WS offer a research approach to link the function of the amygdala and its effects on social behavior.

Abnormal amygdala activity in response to faces has been found in both ASD and WS imaging studies. Hyperactivation of the amygdala was demonstrated when subjects with ASD, as compared to controls, looked at faces¹²³. Furthermore, those with ASD gazed more away than toward the eyes of a presented face, as compared to controls, with a greater amygdala response in subjects with ASD while fixating on the eyes rather than the mouth¹²⁴. This suggests that, in ASD, the amygdala response to faces has a negatively valenced overarousal response. However, other studies showed hypoactivation of the amygdala of subjects with ASD while interpreting emotional states by viewing human eyes¹²⁵ or while processing human fearful faces¹²⁶. In subjects with WS, amygdala reactivity to fearful faces,

presented as negative social stimuli, was drastically attenuated compared with controls¹²⁷, and, strikingly, subjects with WS showed no amygdala activation in a face-discrimination task¹²⁸. The attenuated amygdala activity in WS may result in deficient processing of social-related information, leading subjects to rate strangers' faces as highly approachable⁵⁸. In contrast, higher amygdala reactivity was observed in participants with WS in response to happy faces presented as positive social stimuli¹²⁷.

The common hyperactivation of the amygdala in the two disorders, but in response to opposite stimuli, demonstrates the complexity of amygdala functionality and its relevance to social behavior. Subjects with ASD display aversion-related amygdala activation while eye gazing, resulting in eye contact avoidance. In contrast, the appetitive-related amygdala activation observed in subjects with WS may serve to functionally increase attention to and processing of happy faces. It might be that different subpopulations of neurons, such as glutamatergic or GABAergic, are active in response to the stimuli in these disorders, resulting in the contrasting behavioral phenotypes. Indeed, a recent study showed that in the medial amygdala, a brain region modulating innate social behavior, inhibitory neurons are important in controlling social behavior, while excitatory neurons modulate repetitive social behavior¹²⁹.

When presented with non-social scenes¹³⁰, or threatening scenes but not threatening faces⁹², subjects with WS show increased amygdala activation and abnormal activation of prefrontal regions linked to the amygdala as compared to controls. Indeed, the amygdala–prefrontal circuitry has been shown to be important in the proper representation of the emotional salience of a stimulus (for review, see ref. 131). Normally, the amygdala's output activity is attenuated by the regulation of mPFC excitatory neurons that project and regulate inhibitory neurons in the basolateral amygdala (BLA) or by intercalated cells around the BLA that inhibit output from the central nucleus of the amygdala¹³². Impairments in this circuitry lead to impaired detection of danger, resulting in lower levels of fear and hypersocial behavior, as demonstrated in human and animal models^{133,134}. OFC–amygdala connectivity was functionally disconnected and impaired in subjects with WS⁹², suggesting that impaired prefrontal-regulated inhibition of the amygdala is responsible for the dissociated fear in those subjects, who demonstrate high non-social fear along with low social-related fear. A recent study identified the deficits in the structural integrity of prefrontal–amygdala white matter pathways as the primary cause of this pathology¹³⁵. These findings suggest that increased amygdala activation may play a role in non-social scenarios and the increased generalized anxiety and phobias associated with WS.

The overfriendliness in individuals with WS coexists with non-social anxiety and phobias, suggesting they have lower levels of anxiety that are specific to social stimuli. Indeed, WS and social anxiety disorder (SAD) have multiple opposing characteristics, including general social drive, specific approach to unfamiliar people, social behavior in an unfamiliar social environment, and attention to faces and eye gaze (for review, see ref. 136). Functionally, in subjects with WS, hypoactivation of limbic regions is detected during facial emotion processing when compared to control subjects, while subjects with SAD demonstrate hyperactivation, in addition to hyperactivation in medial frontal regions¹³⁶. This suggests that neural circuits that govern general fear are more functionally separated than those related to social fear and that the latter are oppositely affected in SAD and WS.

Lastly, the amygdala also regulates anxiety, making a simple interpretation of the discussed findings difficult. A direct correlation between anxiety levels and social impairment was observed in the case of ASD¹³⁷, as well as WS¹³⁸. However, in the case of WS, subjects

demonstrate hypersociability along with high anxiety levels. While MRI studies find similar abnormalities in the amygdala in both disorders, the social behavioral phenotypes are opposite, suggesting that subcircuits in either the amygdala or other brain regions upstream or downstream from the amygdala play a role in the opposite social behavior phenotype.

Overall, future studies are needed to better determine the amygdala's valence and function in social behavior, to define the interplay between impaired social behavior and anxiety, and to study whether the different amygdala functions rely on different nuclei that might be oppositely affected in ASD and WS. Since imaging and manipulating the different nuclei of the amygdala is technically difficult in humans, animal models for ASD and WS are valuable research tools for dissecting these questions.

The social motivation theory. The “social motivation theory” suggests that impaired motivation to engage in reciprocal social interaction leads to the ASD-like social deficit¹³⁹. Three key brain regions are related to social motivation and are all highly connected neuroanatomically: orbital and ventromedial regions of the prefrontal cortex, the amygdala, and the ventral striatum. Supporting this theory, children with ASD have a reduced frontostriatal response to socially but not monetarily rewarded learning¹⁴⁰. However, other studies found the deficit in reward processing in subjects with ASD to be attributable not only to social reward, but also to a more general deficit of the reward system¹⁴¹. It is therefore important to determine whether in ASD the impairment is specifically in social motivation or in general motivation, and to study the interplay between the two.

Perhaps one of the most studied molecular mechanisms related to the modulation of social behavior is oxytocin, a neuropeptide synthesized in the hypothalamus, released by the pituitary and affecting the CNS¹⁴². Oxytocin is involved in increasing the degree of approach behavior, social recognition, social memory, the recognition of others' emotions, emotional information processing, maternal behavior and in reducing social fear and anxiety.

Recent studies tested whether oxytocin signaling in mice plays a part in the reward aspect of social interaction¹⁴³. Oxytocin was found to be an enforcement signal in social behavior, acting in medium spiny neurons of the nucleus accumbens (NAc), where it modifies excitatory synaptic transmission by evoking presynaptic long-term depression¹⁴³. Through its abolition in mice, oxytocin was demonstrated to be necessary for social memory, and oxytocin-null mice demonstrate social amnesia that is rescued upon exogenous oxytocin administration¹⁴⁴.

Additional recent studies in mice support the role of reward circuitry in social behavior and show that the ventral tegmental area (VTA), a major source of dopamine in the reward circuitry, is highly active during social interaction¹⁴⁵. Bidirectional control of dopaminergic cells in the VTA modulates social behavior in opposite directions¹⁴⁵. Additionally, activation of the VTA–NAc projection increases social interaction, while VTA–mPFC activation does not affect social interaction, and postsynaptic NAc dopamine receptor D1 medium spiny neurons were shown to be responsible for social behavior regulation¹⁴⁵. Finally, a study on social attachment in monogamous voles showed that dopamine transmission, specifically in the rostral shell of the NAc, promotes pair-bond formation, with D1-like receptor activation decreasing and D2-like receptor activation increasing pair-bond formation¹⁴⁶.

Future directions

Recent development of genome editing techniques such as TALEN¹⁴⁷ and CRISPR¹⁴⁸ will allow us to develop better animal models of

disease, such as primates¹⁴⁹, for social behavioral studies. In particular, the common marmoset, a small New World monkey with rapid reproduction cycles, could contribute to the next generation of genetically engineered models for brain disorder research¹⁵⁰. Common marmosets are small (~350 g), reach sexual maturity at 12–16 months, give birth twice a year, and produce 2–3 offspring with each birth. Marmosets are evolutionarily much closer to humans than rodents in brain structure and function; furthermore, marmosets are very social and communicative and can perform some higher cognitive tasks developed for macaque monkeys. Because of the complexity of genetics in ASD, it would be beneficial to start with monogenic causes of ASD, such as *Shank3* and *Chd8* (chromodomain helicase DNA binding protein 8). For WS, *Gtf2i* would be an excellent candidate for genetic manipulation in marmosets on the basis of knowledge gained from both human and mouse studies. Together, these enabling technologies and new models will likely push the field forward significantly in the next few years.

Note: Any Supplementary Information and Source Data files are available in the online version of the paper.

ACKNOWLEDGMENTS

The authors gratefully acknowledge L. McGrath, F. Dobie, P. Monteiro, A. Krol and Y. Mei for insightful comments on the manuscript. Research in the laboratory of G.F. was supported by the Poitras Center for Affective Disorders Research at MIT, Stanley Center for Psychiatric Research at Broad Institute of MIT and Harvard, National Institute of Mental Health (NIMH), Nancy Lurie Marks Family Foundation, Simons Foundation Autism Research Initiative (SFARI), and Simons Center for the Social Brain at MIT. B.B. was supported by postdoctoral fellowships from the Simons Center for the Social Brain at MIT and from the Autism Science Foundation.

COMPETING FINANCIAL INTERESTS

The authors declare no competing financial interests.

Reprints and permissions information is available online at <http://www.nature.com/reprints/index.html>.

- Frith, C.D. The social brain? *Phil. Trans. R. Soc. Lond. B* **362**, 671–678 (2007).
- Couture, S.M. *et al.* Comparison of social cognitive functioning in schizophrenia and high functioning autism: more convergence than divergence. *Psychol. Med.* **40**, 569–579 (2010).
- Esbensen, A.J., Seltzer, M.M., Lam, K.S. & Bodfish, J.W. Age-related differences in restricted repetitive behaviors in autism spectrum disorders. *J. Autism Dev. Disord.* **39**, 57–66 (2009).
- American Psychiatric Association. *Diagnostic and Statistical Manual of Mental Disorders (DSM-V)* (American Psychiatric Publishing, 2013).
- Kanner, L. Autistic disturbances of affective contact. *Nervous Child* **2**, 217–250 (1943).
- Developmental Disabilities Monitoring Network Surveillance Year 2010 Principal Investigators & Centers for Disease Control and Prevention (CDC). Prevalence of autism spectrum disorder among children aged 8 years — autism and developmental disabilities monitoring network, 11 sites, United States, 2010. *MMWR Surveill. Summ.* **63**, 1–21 (2014).
- Elsabbagh, M. *et al.* Global prevalence of autism and other pervasive developmental disorders. *Autism Res.* **5**, 160–179 (2012).
- Lai, M.-C. *et al.* Cognition in males and females with autism: similarities and differences. *PLoS One* **7**, e47198 (2012).
- Ma, D. *et al.* A genome-wide association study of autism reveals a common novel risk locus at 5p14.1. *Ann. Hum. Genet.* **73**, 263–273 (2009).
- Merikangas, A.K. *et al.* The phenotypic manifestations of rare genic CNVs in autism spectrum disorder. *Mol. Psychiatry* **20**, 1366–1372 (2015).
- Pinto, D. *et al.* Convergence of genes and cellular pathways dysregulated in autism spectrum disorders. *Am. J. Hum. Genet.* **94**, 677–694 (2014).
- Sanders, S.J. *et al.* Insights into autism spectrum disorder genomic architecture and biology from 71 risk loci. *Neuron* **87**, 1215–1233 (2015).
- Weiss, L.A., Arking, D.E., Daly, M.J. & Chakravarti, A. A genome-wide linkage and association scan reveals novel loci for autism. *Nature* **461**, 802–808 (2009).
- Yuen, R.K. *et al.* Whole-genome sequencing of quartet families with autism spectrum disorder. *Nat. Med.* **21**, 185–191 (2015).
- Zoghbi, H.Y. & Bear, M.F. Synaptic dysfunction in neurodevelopmental disorders associated with autism and intellectual disabilities. *Cold Spring Harb. Perspect. Biol.* **4**, pii: a009886 (2012).
- Peça, J. & Feng, G. Cellular and synaptic network defects in autism. *Curr. Opin. Neurobiol.* **22**, 866–872 (2012).
- Sigman, M.D., Kasari, C., Kwon, J.H. & Yirmiya, N. Responses to the negative emotions of others by autistic, mentally retarded, and normal children. *Child Dev.* **63**, 796–807 (1992).
- Lord, C. *et al.* The autism diagnostic observation schedule-generic: a standard measure of social and communication deficits associated with the spectrum of autism. *J. Autism Dev. Disord.* **30**, 205–223 (2000).
- Bauminger, N., Shulman, C. & Agam, G. Peer interaction and loneliness in high-functioning children with autism. *J. Autism Dev. Disord.* **33**, 489–507 (2003).
- Mills, M. & Melhuish, E. Recognition of mother's voice in early infancy. *Nature* **252**, 123–124 (1974).
- Volkmar, F.R. & Mayes, L.C. Gaze behavior in autism. *Dev. Psychopathol.* **2**, 61–69 (1990).
- Mundy, P., Sigman, M. & Kasari, C. A longitudinal study of joint attention and language development in autistic children. *J. Autism Dev. Disord.* **20**, 115–128 (1990).
- Black, M., Freeman, B.J. & Montgomery, J. Systematic observation of play behavior in autistic children. *J. Autism Child. Schizophr.* **5**, 363–371 (1975).
- Williams, J.C., Barratt-Boyes, B.G. & Lowe, J.B. Supravalvular aortic stenosis. *Circulation* **24**, 1311–1318 (1961).
- Mervis, C.B. & John, A.E. Cognitive and behavioral characteristics of children with Williams syndrome: implications for intervention approaches. *Am. J. Med. Genet. C. Semin. Med. Genet.* **154C**, 229–248 (2010).
- Fishman, I., Yam, A., Bellugi, U. & Mills, D. Language and sociability: insights from Williams syndrome **3**, 185–192 (2011).
- Mervis, C.B., Robinson, B.F. & Pani, J.R. Visuospatial construction. *Am. J. Hum. Genet.* **65**, 1222–1229 (1999).
- Dykens, E.M., Rosner, B.A., Ly, T. & Sagun, J. Music and anxiety in Williams syndrome: a harmonious or discordant relationship? *Am. J. Ment. Retard.* **110**, 346–358 (2005).
- Dykens, E.M. Anxiety, fears, and phobias in persons with Williams syndrome. *Dev. Neuropsychol.* **23**, 291–316 (2003).
- Strømme, P., Bjørnstad, P.G. & Ramstad, K. Prevalence estimation of Williams syndrome. *J. Child Neurol.* **17**, 269–271 (2002).
- Pober, B.R. Williams-Beuren syndrome. *N. Engl. J. Med.* **362**, 239–252 (2010).
- Korenberg, J.R. *et al.* VI. Genome structure and cognitive map of Williams syndrome. *J. Cogn. Neurosci.* **12** (suppl. 1), 89–107 (2000).
- Bayés, M., Magano, L.F., Rivera, N., Flores, R. & Pérez Jurado, L.A. Mutational mechanisms of Williams-Beuren syndrome deletions. *Am. J. Hum. Genet.* **73**, 131–151 (2003).
- Edelmann, L. *et al.* An atypical deletion of the Williams-Beuren syndrome interval implicates genes associated with defective visuospatial processing and autism. *J. Med. Genet.* **44**, 136–143 (2007).
- Antonell, A. *et al.* Partial 7q11.23 deletions further implicate GTF2I and GTF2IRD1 as the main genes responsible for the Williams-Beuren syndrome neurocognitive profile. *J. Med. Genet.* **47**, 312–320 (2010).
- Fusco, C. *et al.* Smaller and larger deletions of the Williams Beuren syndrome region implicate genes involved in mild facial phenotype, epilepsy and autistic traits. *Eur. J. Hum. Genet.* **22**, 64–70 (2014).
- Howald, C. *et al.* Two high throughput technologies to detect segmental aneuploidies identify new Williams-Beuren syndrome patients with atypical deletions. *J. Med. Genet.* **43**, 266–273 (2006).
- Beunders, G. *et al.* A triplication of the Williams-Beuren syndrome region in a patient with mental retardation, a severe expressive language delay, behavioural problems and dysmorphisms. *J. Med. Genet.* **47**, 271–275 (2010).
- Depienne, C. *et al.* Autism, language delay and mental retardation in a patient with 7q11 duplication. *J. Med. Genet.* **44**, 452–458 (2007).
- Malenfant, P. *et al.* Association of GTF2I in the Williams-Beuren syndrome critical region with autism spectrum disorders. *J. Autism Dev. Disord.* **42**, 1459–1469 (2012).
- Sanders, S.J. *et al.* Multiple recurrent de novo CNVs, including duplications of the 7q11.23 Williams syndrome region, are strongly associated with autism. *Neuron* **70**, 863–885 (2011).
- Somerville, M.J. *et al.* Severe expressive-language delay related to duplication of the Williams-Beuren locus. *N. Engl. J. Med.* **353**, 1694–1701 (2005).
- Beuren, A.J., Apitz, J. & Harmjan, D. Supravalvular aortic stenosis in association with mental retardation and a certain facial appearance. *Circulation* **26**, 1235–1240 (1962).
- Gosch, A. & Pankau, R. Social-emotional and behavioral adjustment in children with Williams-Beuren syndrome. *Am. J. Med. Genet.* **53**, 335–339 (1994).
- Klein-Tasman, B.P. & Mervis, C.B. Distinctive personality characteristics of 8-, 9-, and 10-year-olds with Williams syndrome. *Dev. Neuropsychol.* **23**, 269–290 (2003).
- Gosch, A. & Pankau, R. Personality characteristics and behaviour problems in individuals of different ages with Williams syndrome. *Dev. Med. Child Neurol.* **39**, 527–533 (1997).
- Riby, D. & Hancock, P.J. Looking at movies and cartoons: eye-tracking evidence from Williams syndrome and autism. *J. Intellect. Disabil. Res.* **53**, 169–181 (2009).
- Dodd, H.F., Porter, M.A., Peters, G.L. & Rapee, R.M. Social approach in pre-school children with Williams syndrome: the role of the face. *J. Intellect. Disabil. Res.* **54**, 194–203 (2010).

49. Mervis, C.B. *et al.* Attentional characteristics of infants and toddlers with Williams syndrome during triadic interactions. *Dev. Neuropsychol.* **23**, 243–268 (2003).
50. Doherty-Sneddon, G., Riby, D.M., Calderwood, L. & Ainsworth, L. Stuck on you: face-to-face arousal and gaze aversion in Williams syndrome. *Cogn. Neuropsychiatry* **14**, 510–523 (2009).
51. Doyle, T.F., Bellugi, U., Korenberg, J.R. & Graham, J. “Everybody in the world is my friend” hypersociability in young children with Williams syndrome. *Am. J. Med. Genet. A* **124A**, 263–273 (2004).
52. Laing, E. *et al.* Atypical development of language and social communication in toddlers with Williams syndrome. *Dev. Sci.* **5**, 233–246 (2002).
53. Elison, S., Stinton, C. & Howlin, P. Health and social outcomes in adults with Williams syndrome: findings from cross-sectional and longitudinal cohorts. *Res. Dev. Disabil.* **31**, 587–599 (2010).
54. Plesa-Skwerer, D., Faja, S., Schofield, C., Verbalis, A. & Tager-Flusberg, H. Perceiving facial and vocal expressions of emotion in individuals with Williams syndrome. *Am. J. Ment. Retard.* **111**, 15–26 (2006).
55. Plesa Skwerer, D. *et al.* A multimeasure approach to investigating affective appraisal of social information in Williams syndrome. *J. Neurodev. Disord.* **3**, 325–334 (2011).
56. Dodd, H.F. & Porter, M.A. I see happy people: attention bias towards happy but not angry facial expressions in Williams syndrome. *Cogn. Neuropsychiatry* **15**, 549–567 (2010).
57. Frigerio, E. *et al.* Is everybody always my friend? Perception of approachability in Williams syndrome. *Neuropsychologia* **44**, 254–259 (2006).
58. Bellugi, U., Adolphs, R., Cassidy, C. & Chiles, M. Towards the neural basis for hypersociability in a genetic syndrome. *Neuroreport* **10**, 1653–1657 (1999).
59. Bayarsaihan, D. *et al.* Genomic organization of the genes *Gtf2ird1*, *Gtf2i*, and *Ncf1* on the mouse chromosome 5 region syntenic to the human chromosome 7q11.23 Williams syndrome critical region. *Genomics* **79**, 137–143 (2002).
60. Botta, A. *et al.* Expression analysis and protein localization of the human *HPC-1/syntaxin 1A*, a gene deleted in Williams syndrome. *Genomics* **62**, 525–528 (1999).
61. Heller, R., Rauch, A., Lüttgen, S., Schröder, B. & Winterpacht, A. Partial deletion of the critical 1.5 Mb interval in Williams-Beuren syndrome. *J. Med. Genet.* **40**, e99 (2003).
62. Morris, C.A. *et al.* GTF2I hemizyosity implicated in mental retardation in Williams syndrome: genotype-phenotype analysis of five families with deletions in the Williams syndrome region. *Am. J. Med. Genet. A* **123A**, 45–59 (2003).
63. van Hagen, J.M. *et al.* Contribution of *CYLN2* and *GTF2IRD1* to neurological and cognitive symptoms in Williams Syndrome. *Neurobiol. Dis.* **26**, 112–124 (2007).
64. Hinsley, T.A., Cunliffe, P., Tipney, H.J., Brass, A. & Tassabehji, M. Comparison of *TFII-I* gene family members deleted in Williams-Beuren syndrome. *Protein Sci.* **13**, 2588–2599 (2004).
65. Roy, A.L. Biochemistry and biology of the inducible multifunctional transcription factor *TFII-I*. *Gene* **274**, 1–13 (2001).
66. Dai, L. *et al.* Is it Williams syndrome? *GTF2IRD1* implicated in visual-spatial construction and *GTF2I* in sociability revealed by high resolution arrays. *Am. J. Med. Genet. A* **149A**, 302–314 (2009).
67. Tassabehji, M. *et al.* *GTF2IRD1* in craniofacial development of humans and mice. *Science* **310**, 1184–1187 (2005).
68. Enkhmandakh, B. *et al.* Essential functions of the Williams-Beuren syndrome-associated *TFII-I* genes in embryonic development. *Proc. Natl. Acad. Sci. USA* **106**, 181–186 (2009).
69. Sakurai, T. *et al.* Haploinsufficiency of *Gtf2i*, a gene deleted in Williams Syndrome, leads to increases in social interactions. *Autism Res.* **4**, 28–39 (2011).
70. Adamo, A. *et al.* 7q11.23 dosage-dependent dysregulation in human pluripotent stem cells affects transcriptional programs in disease-relevant lineages. *Nat. Genet.* **47**, 132–141 (2015).
71. Fletcher, P.C. *et al.* Other minds in the brain: a functional imaging study of “theory of mind” in story comprehension. *Cognition* **57**, 109–128 (1995).
72. Colle, L., Baron-Cohen, S. & Hill, J. Do children with autism have a theory of mind? A non-verbal test of autism vs. specific language impairment. *J. Autism Dev. Disord.* **37**, 716–723 (2007).
73. Embregts, P. & van Nieuwenhuijzen, M. Social information processing in boys with autistic spectrum disorder and mild to borderline intellectual disabilities. *J. Intellect. Disabil. Res.* **53**, 922–931 (2009).
74. Johnson, M.H. Interactive specialization: a domain-general framework for human functional brain development? *Dev. Cogn. Neurosci.* **1**, 7–21 (2011).
75. Nelson, S.B. & Valakh, V. Excitatory/inhibitory balance and circuit homeostasis in autism spectrum disorders. *Neuron* **87**, 684–698 (2015).
76. Leekam, S.R., Nieto, C., Libby, S.J., Wing, L. & Gould, J. Describing the sensory abnormalities of children and adults with autism. *J. Autism Dev. Disord.* **37**, 894–910 (2007).
77. Yirmiya, N. & Charman, T. The prodrome of autism: early behavioral and biological signs, regression, peri- and post-natal development and genetics. *J. Child Psychol. Psychiatry* **51**, 432–458 (2010).
78. Jones, E.J., Gliga, T., Bedford, R., Charman, T. & Johnson, M.H. Developmental pathways to autism: a review of prospective studies of infants at risk. *Neurosci. Biobehav. Rev.* **39**, 1–33 (2014).
79. Wolff, J.J. *et al.* Altered corpus callosum morphology associated with autism over the first 2 years of life. *Brain* **138**, 2046–2058 (2015).
80. Wolff, J.J. *et al.* Differences in white matter fiber tract development present from 6 to 24 months in infants with autism. *Am. J. Psychiatry* **169**, 589–600 (2012).
81. Hazlett, H.C. *et al.* Early brain overgrowth in autism associated with an increase in cortical surface area before age 2 years. *Arch. Gen. Psychiatry* **68**, 467–476 (2011).
82. Schumann, C.M. *et al.* Longitudinal magnetic resonance imaging study of cortical development through early childhood in autism. *J. Neurosci.* **30**, 4419–4427 (2010).
83. Hazlett, H.C. *et al.* Brain volume findings in 6-month-old infants at high familial risk for autism. *Am. J. Psychiatry* **169**, 601–608 (2012).
84. Carper, R.A., Moses, P., Tighe, Z.D. & Courchesne, E. Cerebral lobes in autism: early hyperplasia and abnormal age effects. *Neuroimage* **16**, 1038–1051 (2002).
85. Murdaugh, D.L. *et al.* Differential deactivation during mentalizing and classification of autism based on default mode network connectivity. *PLoS One* **7**, e50064 (2012).
86. Redcay, E. *et al.* Atypical brain activation patterns during a face-to-face joint attention game in adults with autism spectrum disorder. *Hum. Brain Mapp.* **34**, 2511–2523 (2013).
87. Travers, B.G. *et al.* Diffusion tensor imaging in autism spectrum disorder: a review. *Autism Res.* **5**, 289–313 (2012).
88. Dajani, D.R. & Uddin, L.Q. Local brain connectivity across development in autism spectrum disorder: A cross-sectional investigation. *Autism Res.* **9**, 43–54 (2016).
89. Venkataraman, A., Duncan, J.S., Yang, D.Y. & Pelphrey, K.A. An unbiased Bayesian approach to functional connectomics implicates social-communication networks in autism. *Neuroimage Clin.* **8**, 356–366 (2015).
90. Bailey, A. *et al.* A clinicopathological study of autism. *Brain* **121**, 889–905 (1998).
91. Buxhoeveden, D.P. *et al.* Reduced minicolumns in the frontal cortex of patients with autism. *Neuropathol. Appl. Neurobiol.* **32**, 483–491 (2006).
92. Meyer-Lindenberg, A. *et al.* Neural correlates of genetically abnormal social cognition in Williams syndrome. *Nat. Neurosci.* **8**, 991–993 (2005).
93. Mobbs, D. *et al.* Frontostriatal dysfunction during response inhibition in Williams syndrome. *Biol. Psychiatry* **62**, 256–261 (2007).
94. Porter, M.A., Coltheart, M. & Langdon, R. The neuropsychological basis of hypersociability in Williams and Down syndrome. *Neuropsychologia* **45**, 2839–2849 (2007).
95. Little, K. *et al.* Heterogeneity of social approach behaviour in Williams syndrome: the role of response inhibition. *Res. Dev. Disabil.* **34**, 959–967 (2013).
96. Mimura, M. *et al.* A preliminary study of orbitofrontal activation and hypersociability in Williams syndrome. *J. Neurodev. Disord.* **2**, 93–98 (2010).
97. Baroncelli, L. *et al.* Brain plasticity and disease: a matter of inhibition. *Neural Plast.* **2011**, 286073 (2011).
98. Gogolla, N., Takesian, A.E., Feng, G., Fagioli, M. & Hensch, T.K. Sensory integration in mouse insular cortex reflects GABA circuit maturation. *Neuron* **83**, 894–905 (2014).
99. Peça, J. *et al.* *Shank3* mutant mice display autistic-like behaviours and striatal dysfunction. *Nature* **472**, 437–442 (2011).
100. Yizhar, O. *et al.* Neocortical excitation/inhibition balance in information processing and social dysfunction. *Nature* **477**, 171–178 (2011).
101. Lee, J. *et al.* *Shank3*-mutant mice lacking exon 9 show altered excitation/inhibition balance, enhanced rearing, and spatial memory deficit. *Front. Cell. Neurosci.* **9**, 94 (2015).
102. Kim, E. *et al.* GKAP, a novel synaptic protein that interacts with the guanylate kinase-like domain of the PSD-95/SAP90 family of channel clustering molecules. *J. Cell Biol.* **136**, 669–678 (1997).
103. Boeckers, T.M., Bockmann, J., Kreutz, M.R. & Gundelfinger, E.D. ProSAP/Shank proteins – a family of higher order organizing molecules of the postsynaptic density with an emerging role in human neurological disease. *J. Neurochem.* **81**, 903–910 (2002).
104. Takeuchi, M. *et al.* SAPAPs. A family of PSD-95/SAP90-associated proteins localized at postsynaptic density. *J. Biol. Chem.* **272**, 11943–11951 (1997).
105. Bozdagi, O. *et al.* Haploinsufficiency of the autism-associated *Shank3* gene leads to deficits in synaptic function, social interaction, and social communication. *Mol. Autism* **1**, 15 (2010).
106. Schmeisser, M.J. *et al.* Autistic-like behaviours and hyperactivity in mice lacking ProSAP1/Shank2. *Nature* **486**, 256–260 (2012).
107. Wang, X. *et al.* Synaptic dysfunction and abnormal behaviors in mice lacking major isoforms of *Shank3*. *Hum. Mol. Genet.* **20**, 3093–3108 (2011).
108. Won, H. *et al.* Autistic-like social behaviour in *Shank2*-mutant mice improved by restoring NMDA receptor function. *Nature* **486**, 261–265 (2012).
109. Yang, M. *et al.* Reduced excitatory neurotransmission and mild autism-relevant phenotypes in adolescent *Shank3* null mutant mice. *J. Neurosci.* **32**, 6525–6541 (2012).
110. Zhou, Y. *et al.* Mice with *Shank3* Mutations associated with ASD and schizophrenia display both shared and distinct defects. *Neuron* **89**, 147–162 (2016).
111. Banerjee, A., Castro, J. & Sur, M. Rett syndrome: genes, synapses, circuits, and therapeutics. *Front. Psychiatry* **3**, 34 (2012).
112. Chao, H.T. *et al.* Dysfunction in GABA signaling mediates autism-like stereotypies and Rett syndrome phenotypes. *Nature* **468**, 263–269 (2010).

113. Silverman, J.L. *et al.* GABAB receptor agonist R-baclofen reverses social deficits and reduces repetitive behavior in two mouse models of autism. *Neuropsychopharmacology* **40**, 2228–2239 (2015).
114. Cellot, G. & Cherubini, E. Reduced inhibitory gate in the barrel cortex of Neuroigin3R451C knock-in mice, an animal model of autism spectrum disorders. *Physiol. Rep.* **2**, pii: e12077 (2014).
115. Fried, I., MacDonald, K.A. & Wilson, C.L. Single neuron activity in human hippocampus and amygdala during recognition of faces and objects. *Neuron* **18**, 753–765 (1997).
116. Spezio, M.L., Huang, P.Y., Castelli, F. & Adolphs, R. Amygdala damage impairs eye contact during conversations with real people. *J. Neurosci.* **27**, 3994–3997 (2007).
117. Adolphs, R., Sears, L. & Piven, J. Abnormal processing of social information from faces in autism. *J. Cogn. Neurosci.* **13**, 232–240 (2001).
118. Adolphs, R., Tranel, D. & Damasio, A.R. The human amygdala in social judgment. *Nature* **393**, 470–474 (1998).
119. Sturm, V. *et al.* DBS in the basolateral amygdala improves symptoms of autism and related self-injurious behavior: a case report and hypothesis on the pathogenesis of the disorder. *Front. Hum. Neurosci.* **6**, 341 (2012).
120. Schumann, C.M. *et al.* The amygdala is enlarged in children but not adolescents with autism; the hippocampus is enlarged at all ages. *J. Neurosci.* **24**, 6392–6401 (2004).
121. Haas, B.W., Sheau, K., Kelley, R.G., Thompson, P.M. & Reiss, A.L. Regionally specific increased volume of the amygdala in Williams syndrome: Evidence from surface-based modeling. *Hum. Brain Mapp.* **35**, 866–874 (2014).
122. Martens, M.A., Wilson, S.J., Dudgeon, P. & Reutens, D.C. Approachability and the amygdala: insights from Williams syndrome. *Neuropsychologia* **47**, 2446–2453 (2009).
123. Dalton, K.M. *et al.* Gaze fixation and the neural circuitry of face processing in autism. *Nat. Neurosci.* **8**, 519–526 (2005).
124. Kliemann, D., Dziobek, I., Hatri, A., Baudewig, J. & Heekeren, H.R. The role of the amygdala in atypical gaze on emotional faces in autism spectrum disorders. *J. Neurosci.* **32**, 9469–9476 (2012).
125. Baron-Cohen, S. *et al.* Social intelligence in the normal and autistic brain: an fMRI study. *Eur. J. Neurosci.* **11**, 1891–1898 (1999).
126. Ashwin, C., Baron-Cohen, S., Wheelwright, S., O'Riordan, M. & Bullmore, E.T. Differential activation of the amygdala and the 'social brain' during fearful face-processing in Asperger Syndrome. *Neuropsychologia* **45**, 2–14 (2007).
127. Haas, B.W. *et al.* Genetic influences on sociability: heightened amygdala reactivity and event-related responses to positive social stimuli in Williams syndrome. *J. Neurosci.* **29**, 1132–1139 (2009).
128. Paul, B.M. *et al.* Amygdala response to faces parallels social behavior in Williams syndrome. *Soc. Cogn. Affect. Neurosci.* **4**, 278–285 (2009).
129. Hong, W., Kim, D.W. & Anderson, D.J. Antagonistic control of social versus repetitive self-grooming behaviors by separable amygdala neuronal subsets. *Cell* **158**, 1348–1361 (2014).
130. Muñoz, K.E. *et al.* Abnormalities in neural processing of emotional stimuli in Williams syndrome vary according to social vs. non-social content. *Neuroimage* **50**, 340–346 (2010).
131. Bishop, S.J. Neurocognitive mechanisms of anxiety: an integrative account. *Trends Cogn. Sci.* **11**, 307–316 (2007).
132. Quirk, G.J., Likhtik, E., Pelletier, J.G. & Paré, D. Stimulation of medial prefrontal cortex decreases the responsiveness of central amygdala output neurons. *J. Neurosci.* **23**, 8800–8807 (2003).
133. Amaral, D.G. The amygdala, social behavior, and danger detection. *Ann. NY Acad. Sci.* **1000**, 337–347 (2003).
134. Machado, C.J., Kazama, A.M. & Bachevalier, J. Impact of amygdala, orbital frontal, or hippocampal lesions on threat avoidance and emotional reactivity in nonhuman primates. *Emotion* **9**, 147–163 (2009).
135. Avery, S.N., Thornton-Wells, T.A., Anderson, A.W. & Blackford, J.U. White matter integrity deficits in prefrontal-amygdala pathways in Williams syndrome. *Neuroimage* **59**, 887–894 (2012).
136. Binelli, C. *et al.* Common and distinct neural correlates of facial emotion processing in social anxiety disorder and Williams syndrome: a systematic review and voxel-based meta-analysis of functional resonance imaging studies. *Neuropsychologia* **64C**, 205–217 (2014).
137. White, S.W., Oswald, D., Ollendick, T. & Scahill, L. Anxiety in children and adolescents with autism spectrum disorders. *Clin. Psychol. Rev.* **29**, 216–229 (2009).
138. Riby, D.M. *et al.* The interplay between anxiety and social functioning in Williams syndrome. *J. Autism Dev. Disord.* **44**, 1220–1229 (2014).
139. Chevallier, C., Kohls, G., Troiani, V., Brodtkin, E.S. & Schultz, R.T. The social motivation theory of autism. *Trends Cogn. Sci.* **16**, 231–239 (2012).
140. Scott-Van Zeeland, A.A., Dapretto, M., Ghahremani, D.G., Poldrack, R.A. & Bookheimer, S.Y. Reward processing in autism. *Autism Res.* **3**, 53–67 (2010).
141. Kohls, G. *et al.* Atypical brain responses to reward cues in autism as revealed by event-related potentials. *J. Autism Dev. Disord.* **41**, 1523–1533 (2011).
142. Heinrichs, M., von Dawans, B. & Domes, G. Oxytocin, vasopressin, and human social behavior. *Front. Neuroendocrinol.* **30**, 548–557 (2009).
143. Dölen, G., Darvishzadeh, A., Huang, K.W. & Malenka, R.C. Social reward requires coordinated activity of nucleus accumbens oxytocin and serotonin. *Nature* **501**, 179–184 (2013).
144. Ferguson, J.N. *et al.* Social amnesia in mice lacking the oxytocin gene. *Nat. Genet.* **25**, 284–288 (2000).
145. Gunaydin, L.A. *et al.* Natural neural projection dynamics underlying social behavior. *Cell* **157**, 1535–1551 (2014).
146. Aragona, B.J. *et al.* Nucleus accumbens dopamine differentially mediates the formation and maintenance of monogamous pair bonds. *Nat. Neurosci.* **9**, 133–139 (2006).
147. Zhang, F. *et al.* Efficient construction of sequence-specific TAL effectors for modulating mammalian transcription. *Nat. Biotechnol.* **29**, 149–153 (2011).
148. Cong, L. *et al.* Multiplex genome engineering using CRISPR/Cas systems. *Science* **339**, 819–823 (2013).
149. Niu, Y. *et al.* Generation of gene-modified cynomolgus monkey via Cas9/RNA-mediated gene targeting in one-cell embryos. *Cell* **156**, 836–843 (2014).
150. Okano, H., Hikishima, K., Iriki, A. & Sasaki, E. The common marmoset as a novel animal model system for biomedical and neuroscience research applications. *Semin. Fetal Neonatal Med.* **17**, 336–340 (2012).